

Transcriptomics

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Outline

Introduction to sequencing technologies

- High throughput sequencing
- Overview of the sequencing platforms

Experimental design

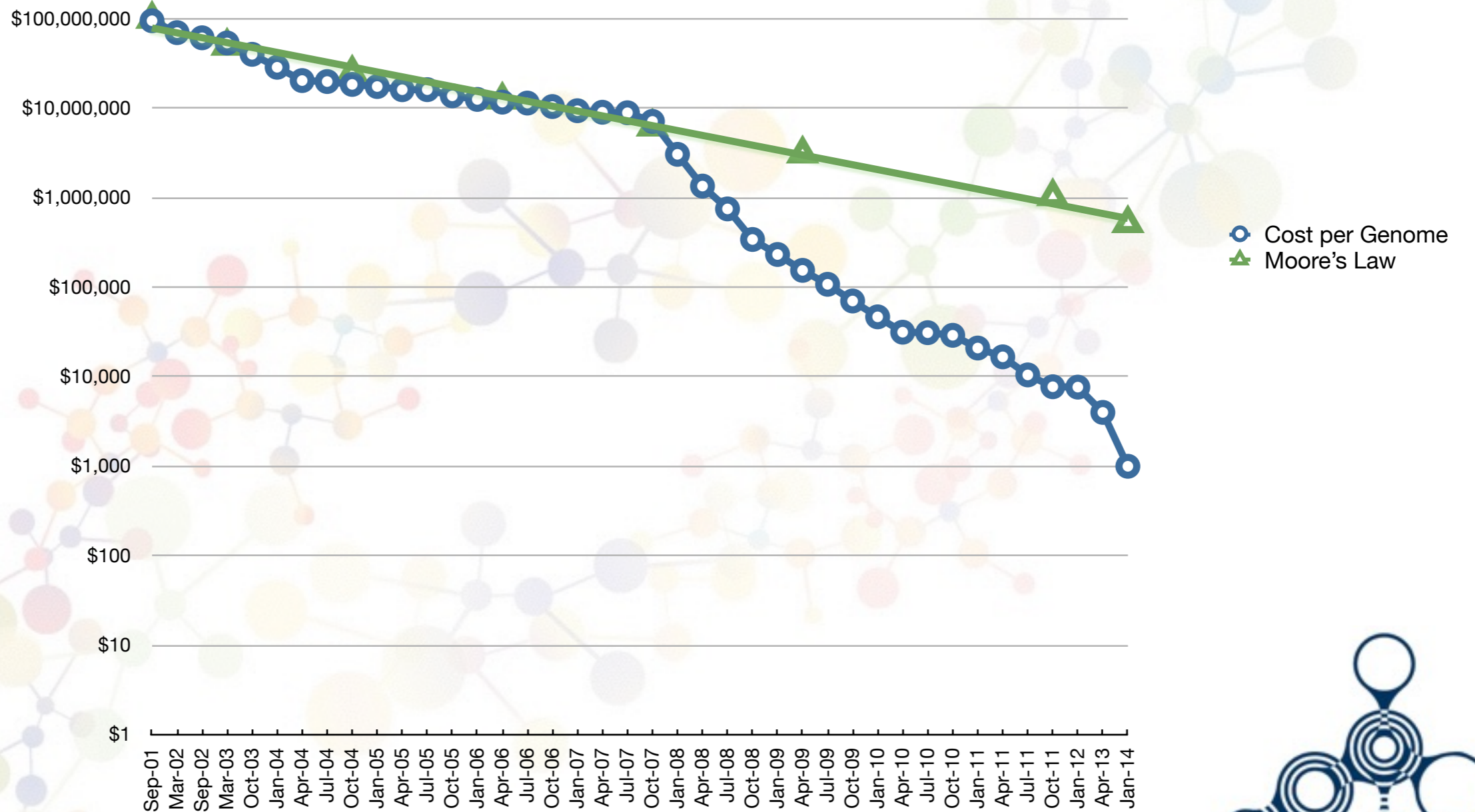
Analysis of transcriptomics data

Case studies

- Drosophila study
- No genome available



Cost of sequencing

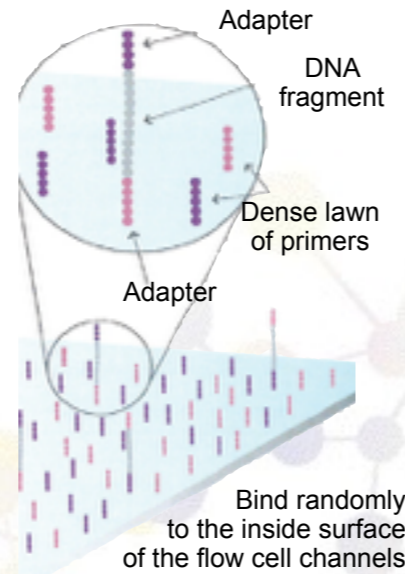


Illumina sequencing

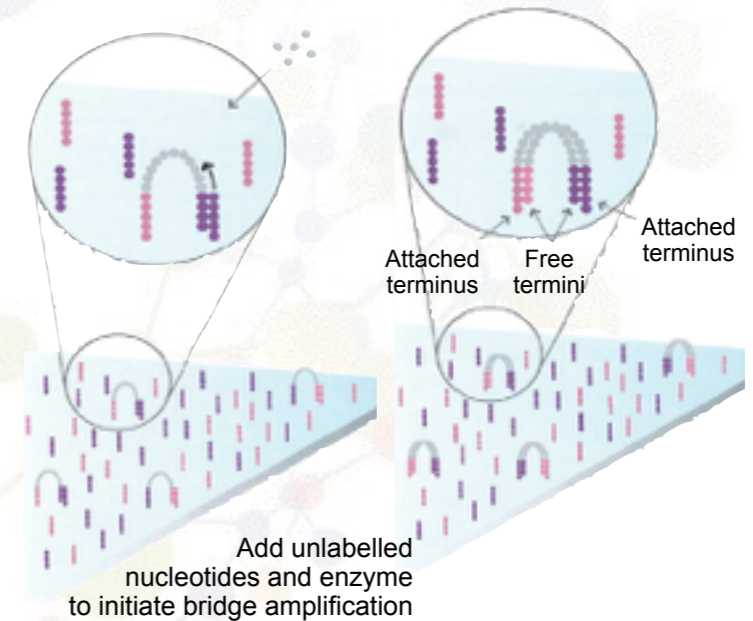
- Cluster generation



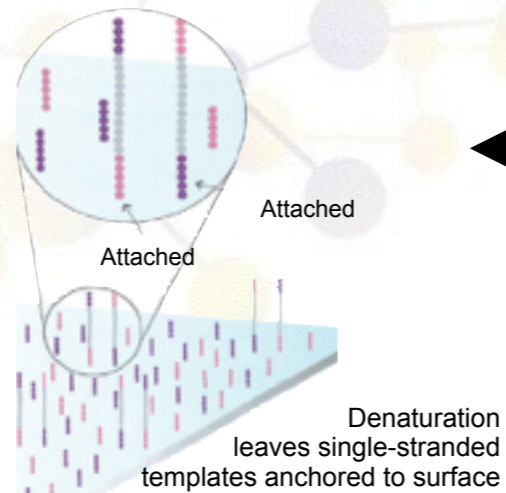
Attach DNA to surface



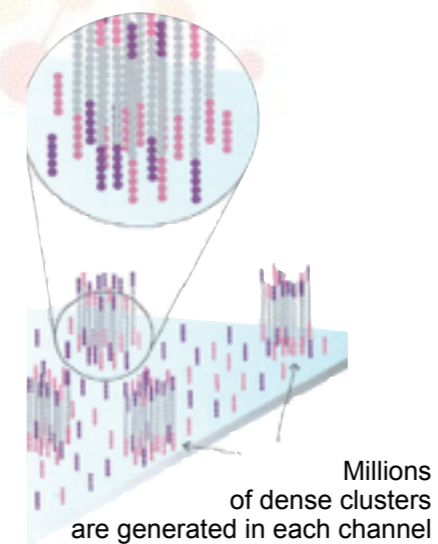
Bridge amplification



Denature ds molecules



Complete amplification

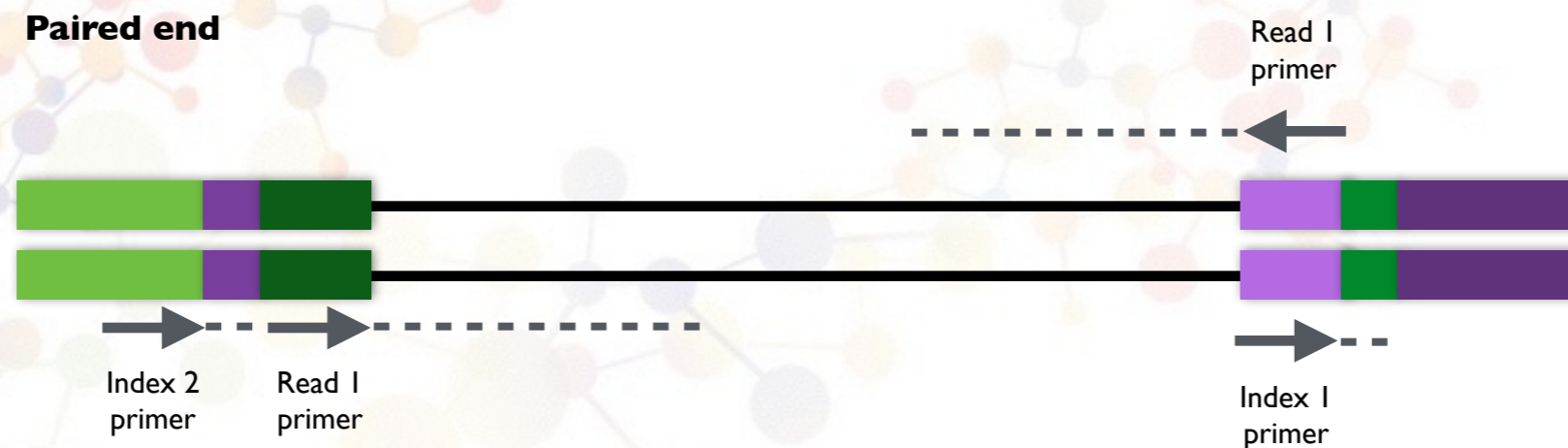


Paired vs Single end sequencing

Single end



Paired end



Illumina

- Base by base sequencing
- Reads all the same length
- Short reads
- High quality

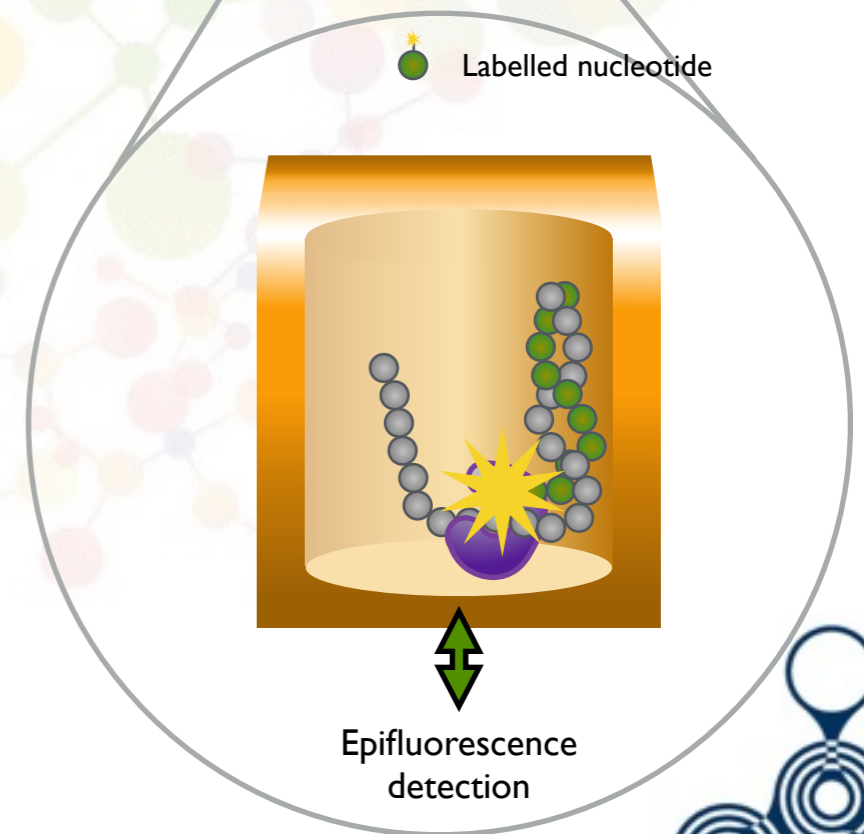
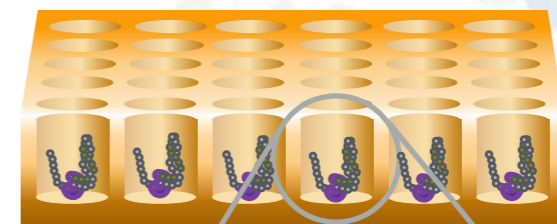


Pacific Biosciences

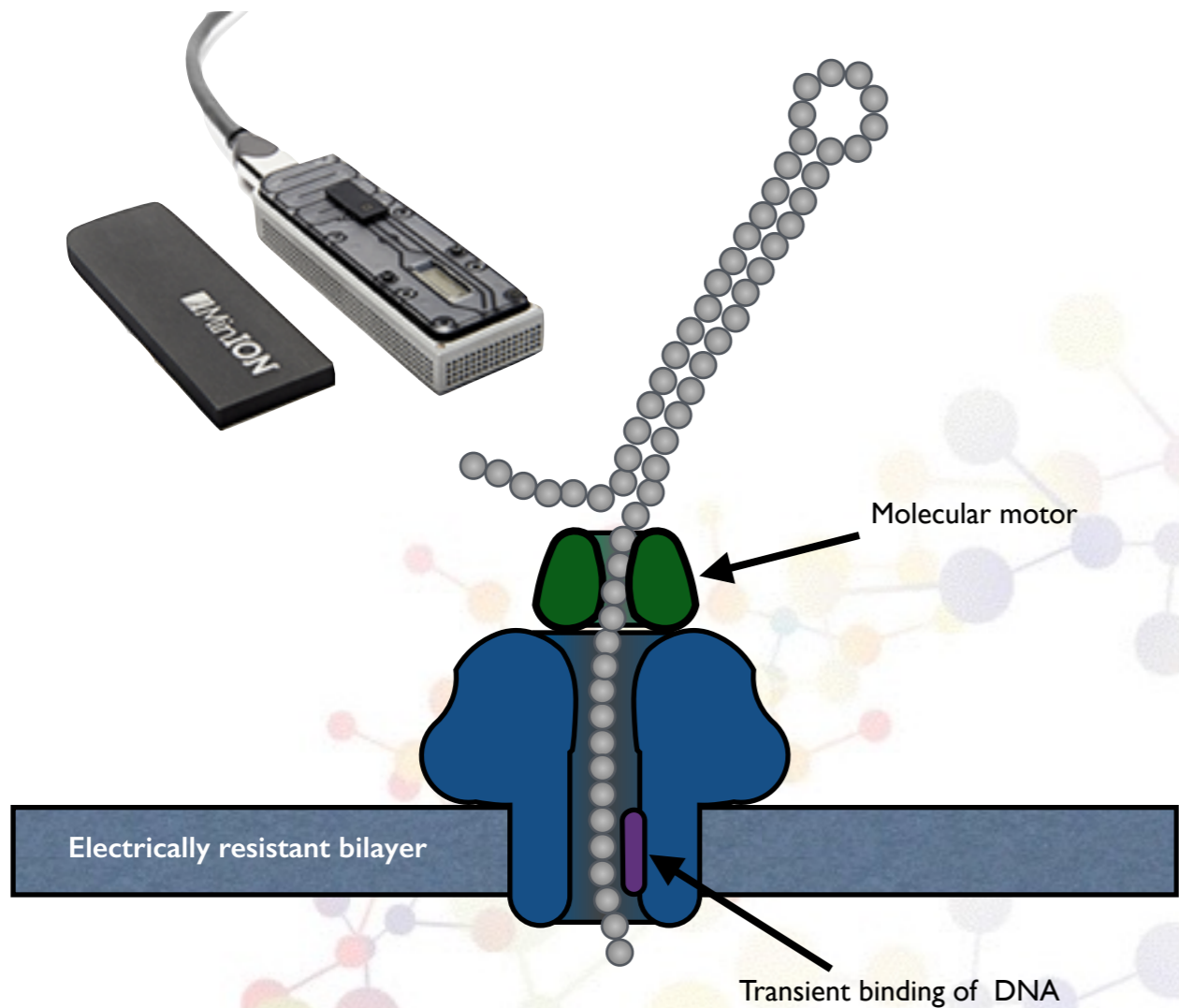
Real time single molecule sequencing

Long reads up to 20kb

Direct sequencing of modified bases e.g. methylated Cytosine



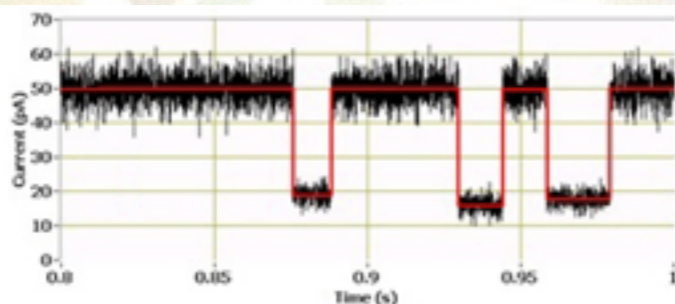
Oxford Nanopore



Real time single molecule sequencing

Long read lengths
10's of kilo bases

Can detect modified DNA bases



Many applications

Total RNA

micro RNA

Immuno precipitated RNA



Template preparation & Sequencing



Data processing, base calling & storage

Mapping to reference genome

De novo assembly

Transcript differential expression

Novel transcript discovery

Differential isoform usage

micro RNA analysis

RIP-Seq analysis



Experimental design

- Key to an expression study
- Ensure that the questions can be answered within experimental constraints
 - Cost
 - Availability of RNA

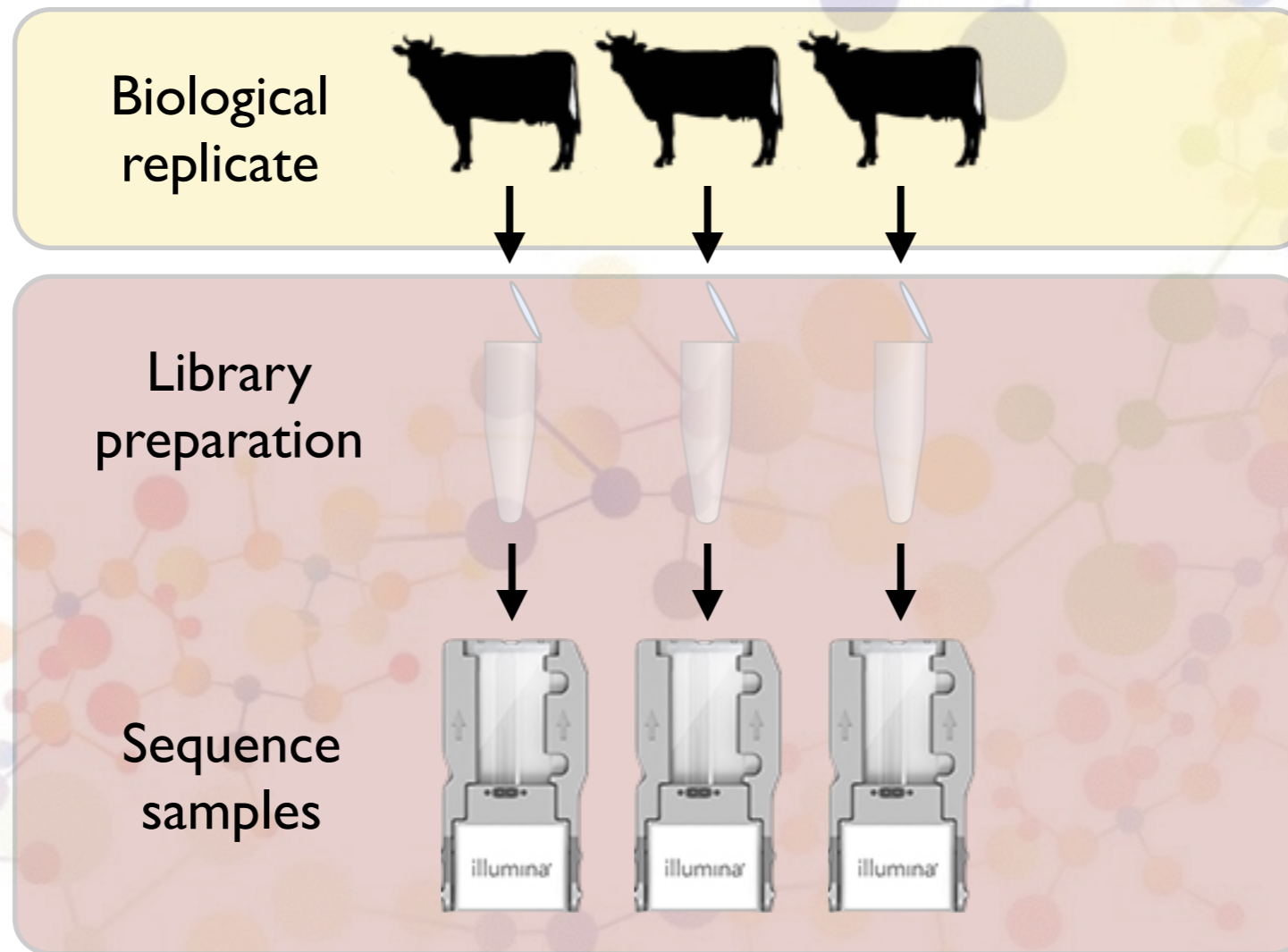


Variation in RNA-Seq experiments

- Biological variation
- Technical variation



Levels of variation



Flowcell
effect

<

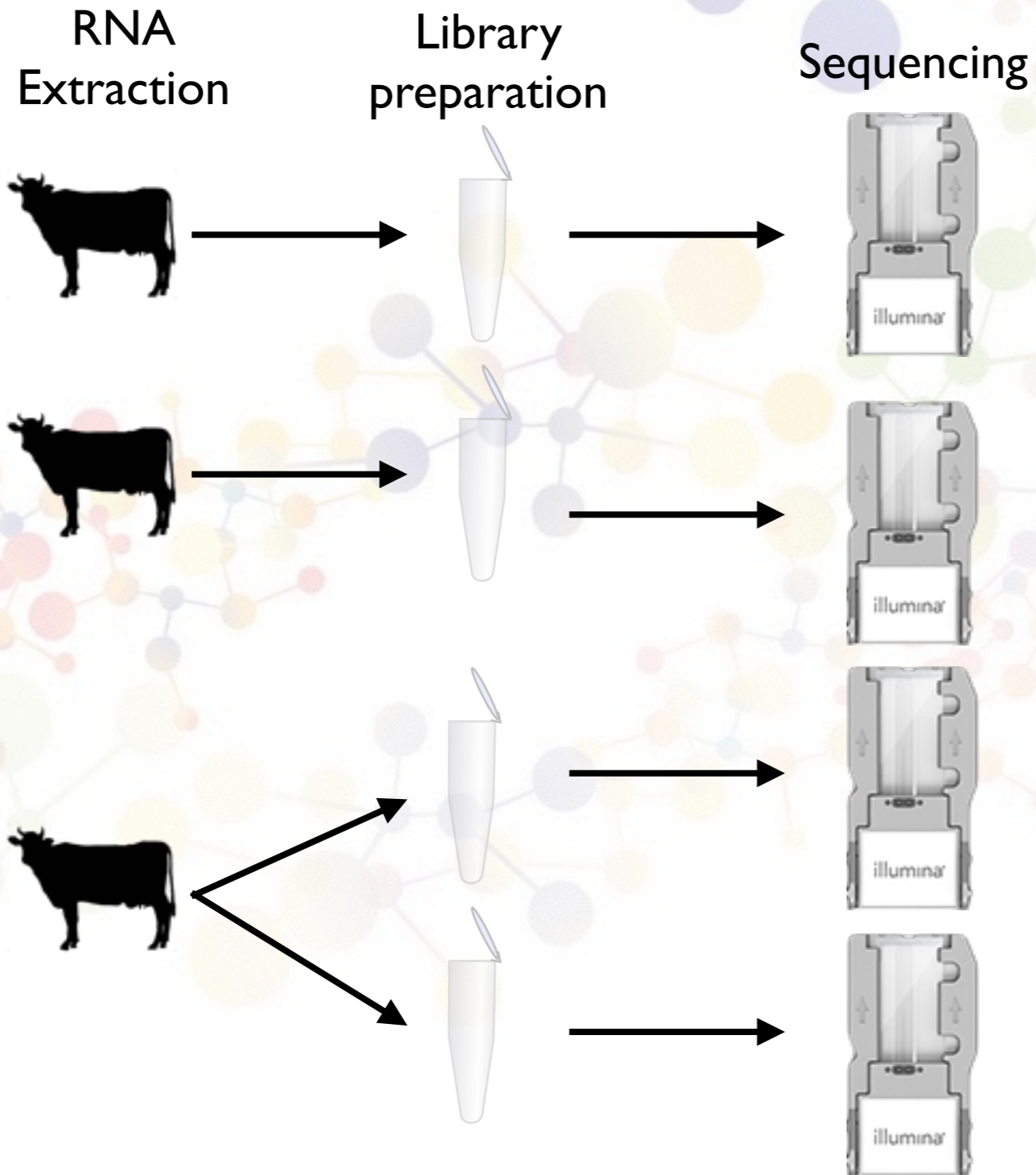
Library prep
effect

<<

Biological
effect



Replicates



Replicates

- Biological replication is essential
- How many replicates?
- Technical variation
- Barcode and multiplex



Sequencing depth

- Difficult to predict
- Estimates can be based on the length of the transcriptome and the expected dynamic range of transcript abundances
- Greater sequence depth increases sensitivity to detect smaller changes and lower expressed transcripts



**What do I need to
provide?**



Poly A selection

- 1 μg total RNA
- Prepare using tri reagent or Qiagen Total RNA kit
- Recommend paired end sequencing



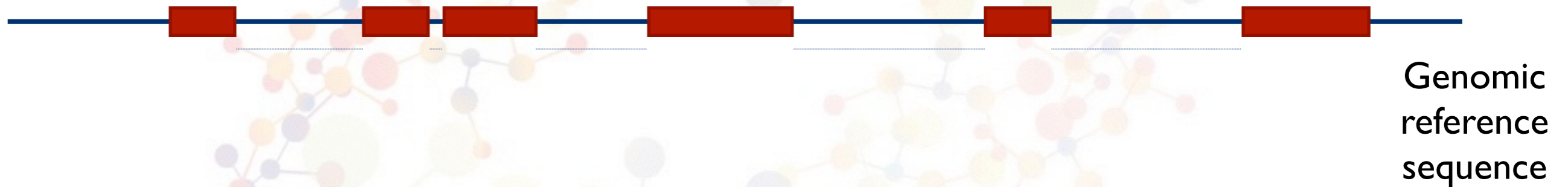
Ribosomal reduction

- 10ng to 400ng ribosomal reduced RNA
- Ribosomal reduction kits from Qiagen, Illumina, Thermo, NEB etc
- Needs optimisation
- Prepare using tri reagent or Qiagen Total RNA kit
- Recommend paired end sequencing
- More reads

Alignment



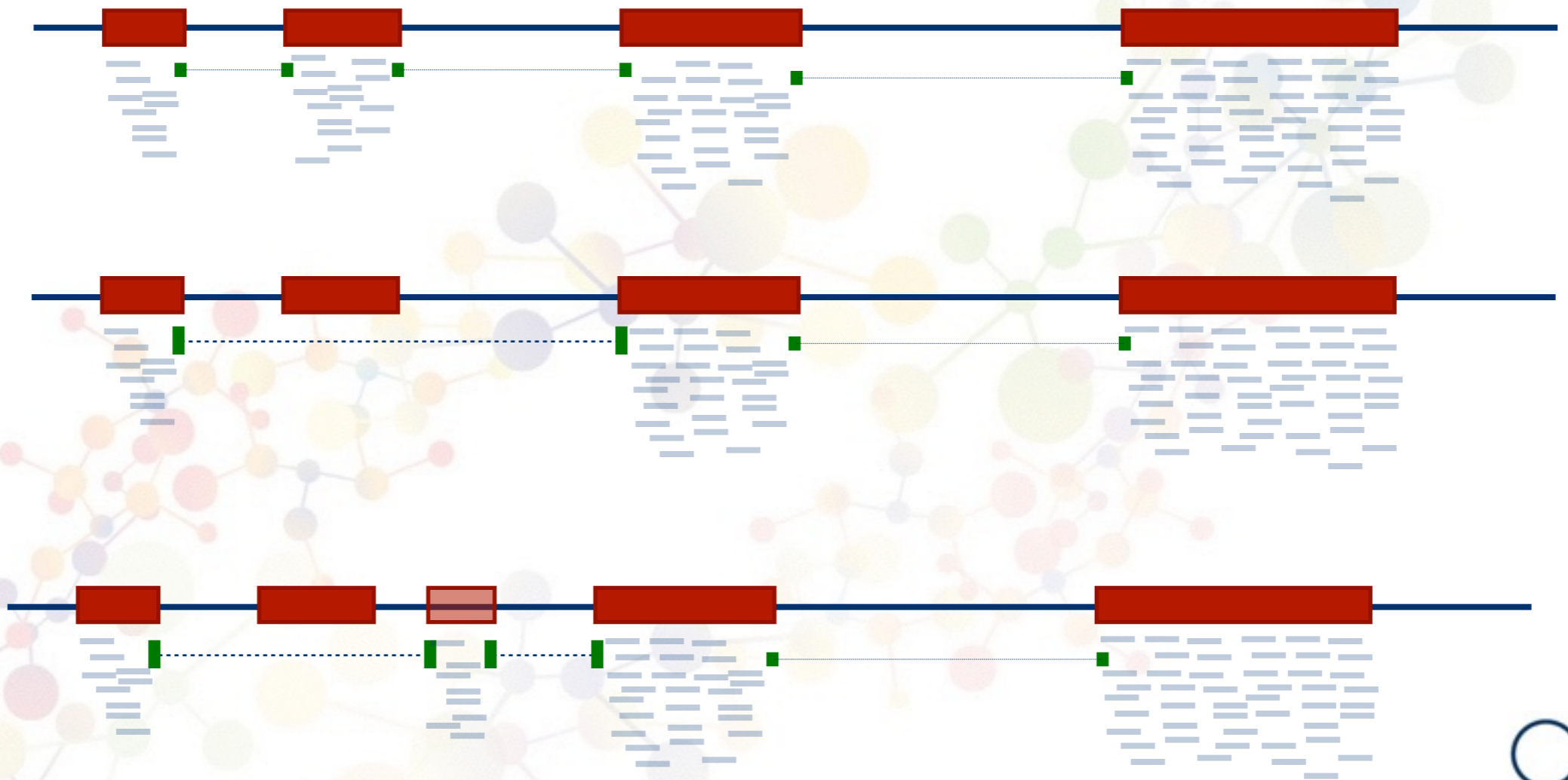
Alignment-Tophat



Alignment-Tophat

Exon skipped

Novel exon

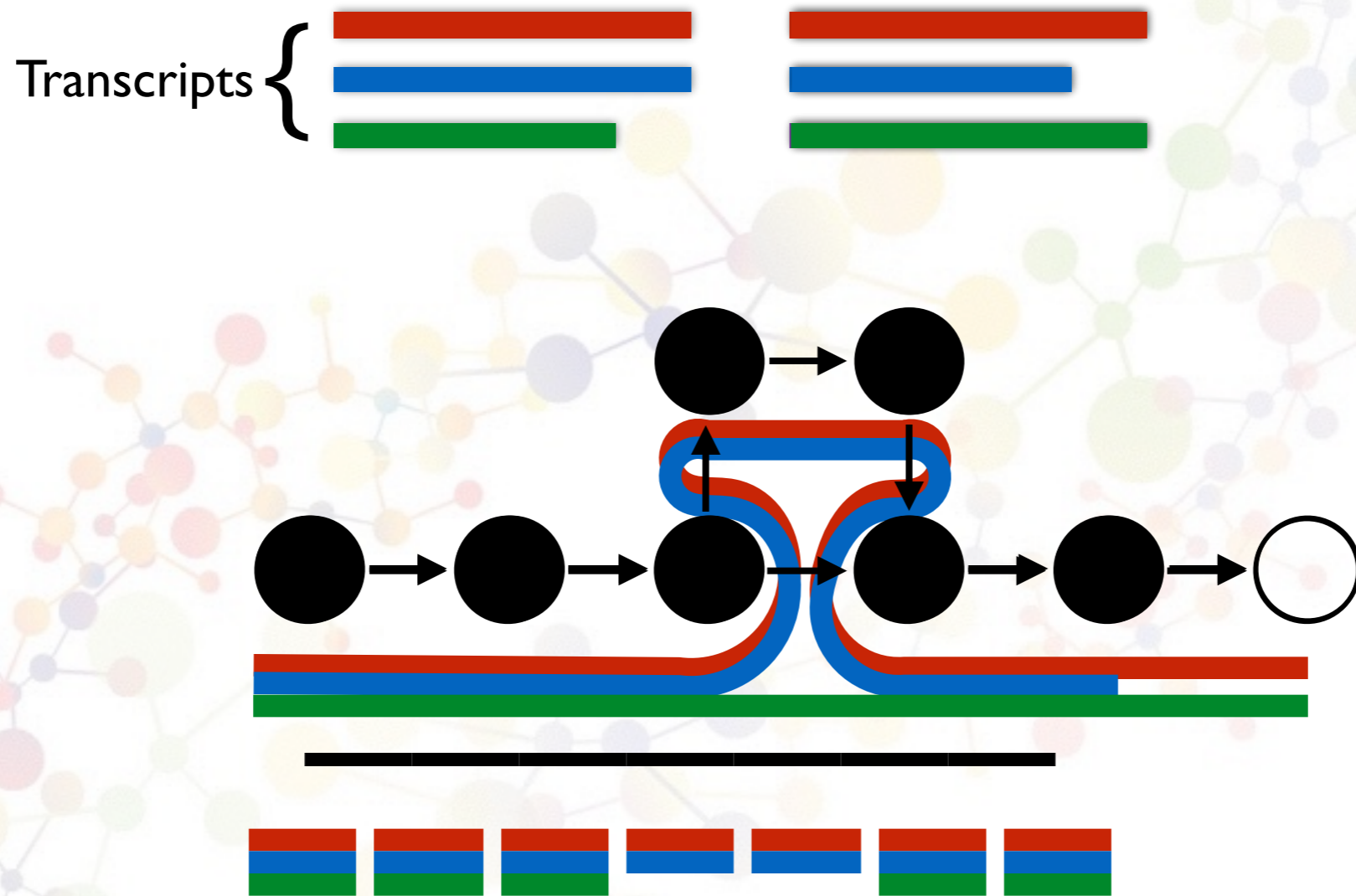


HISAT

- Similar to Tophat
- Two types of index
- An index of the whole genome
- Multiple indexes of overlapping ~64kb chunks of the genome



Kallisto



Differential expression

- Several sources of variability
- Batch effects
- Over dispersion
- Uncertainty in count origin



Software tools

Cufflinks - Differential analysis at transcript level

DESeq2 - Pools information across genes.
Shrinkage estimation for dispersion and fold changes

Voom - Transforms transcript count data to logCPM and maps each observation to the mean variance

Novel insecticide target discovery by RNA Sequencing



Background

Insects dominant life form

Disease vectors

- malaria, yellow fever, sleeping sickness

15-25% loss to GDP worldwide

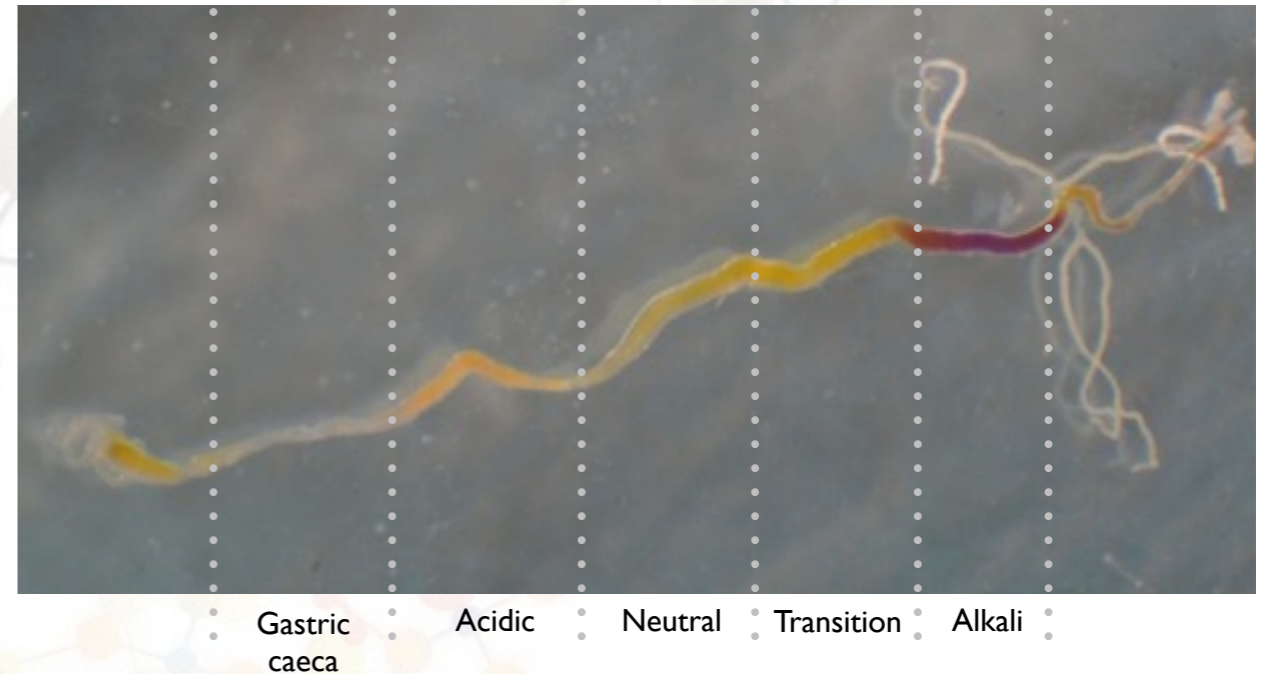
Few new pesticides

Must not harm pollinators

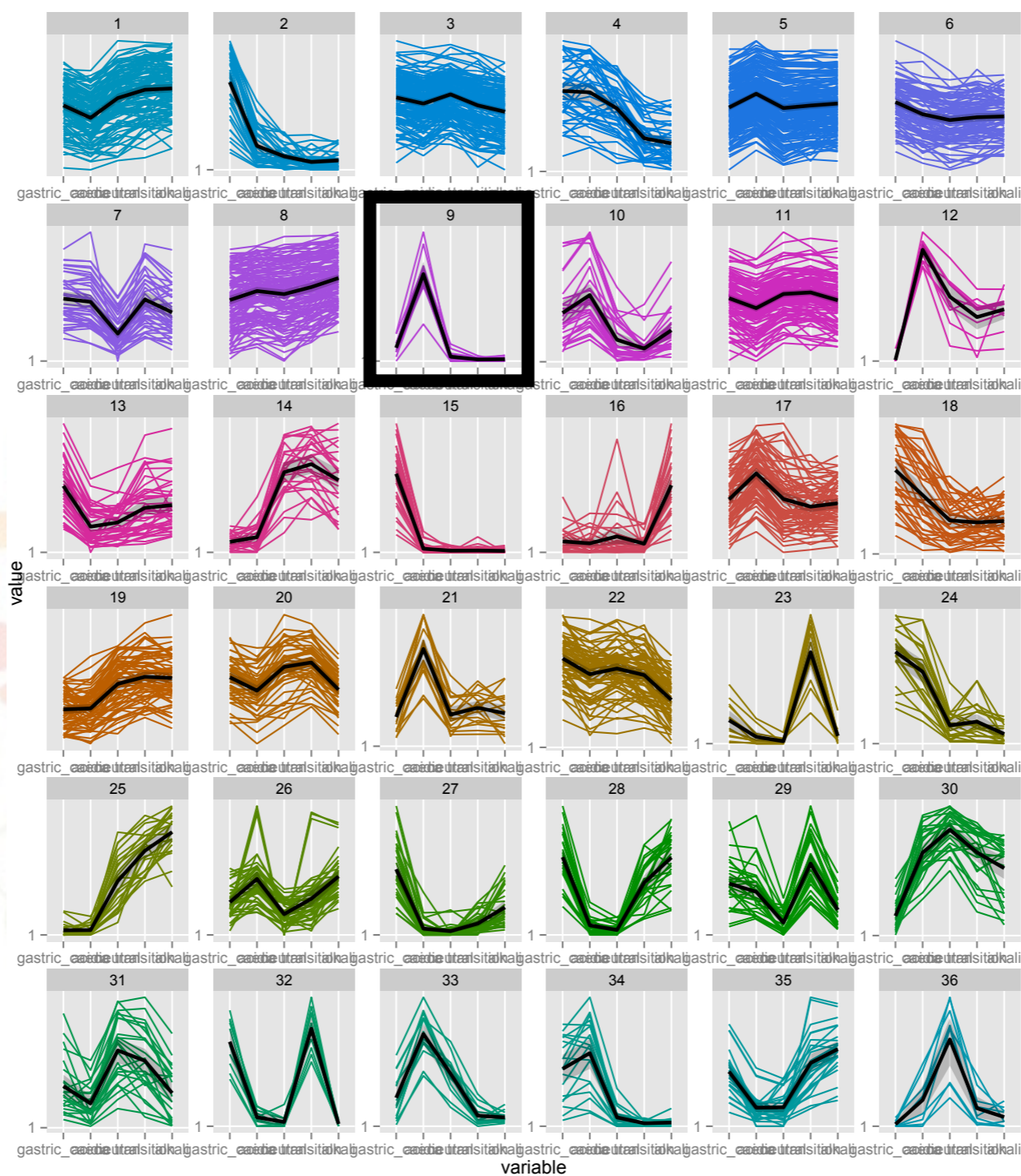


Transcriptomics

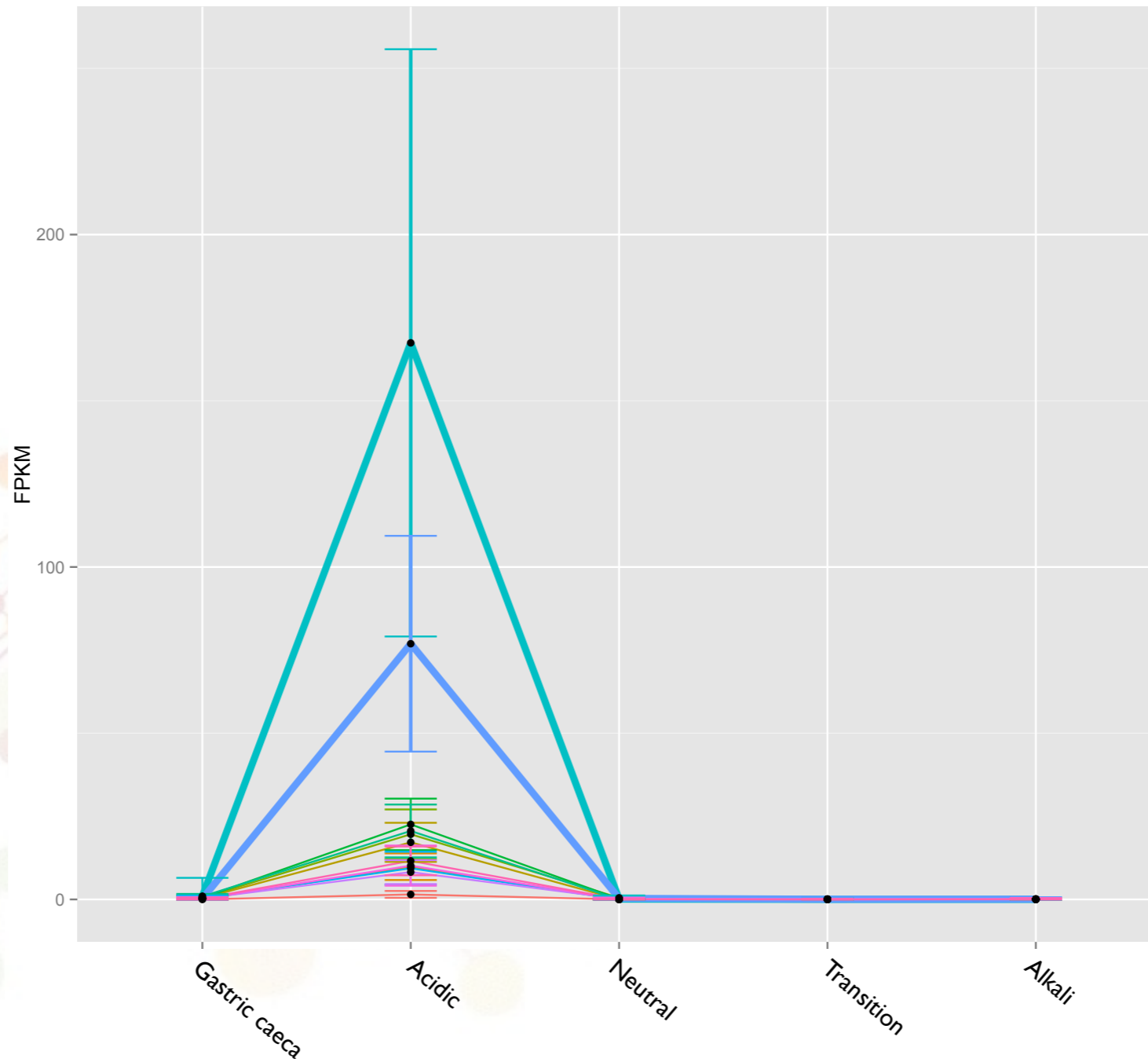
- RNA isolated from 5 distinct regions of the digestive tract
- Samples were sequenced
- Compare sample to discover differently expressed transcripts.



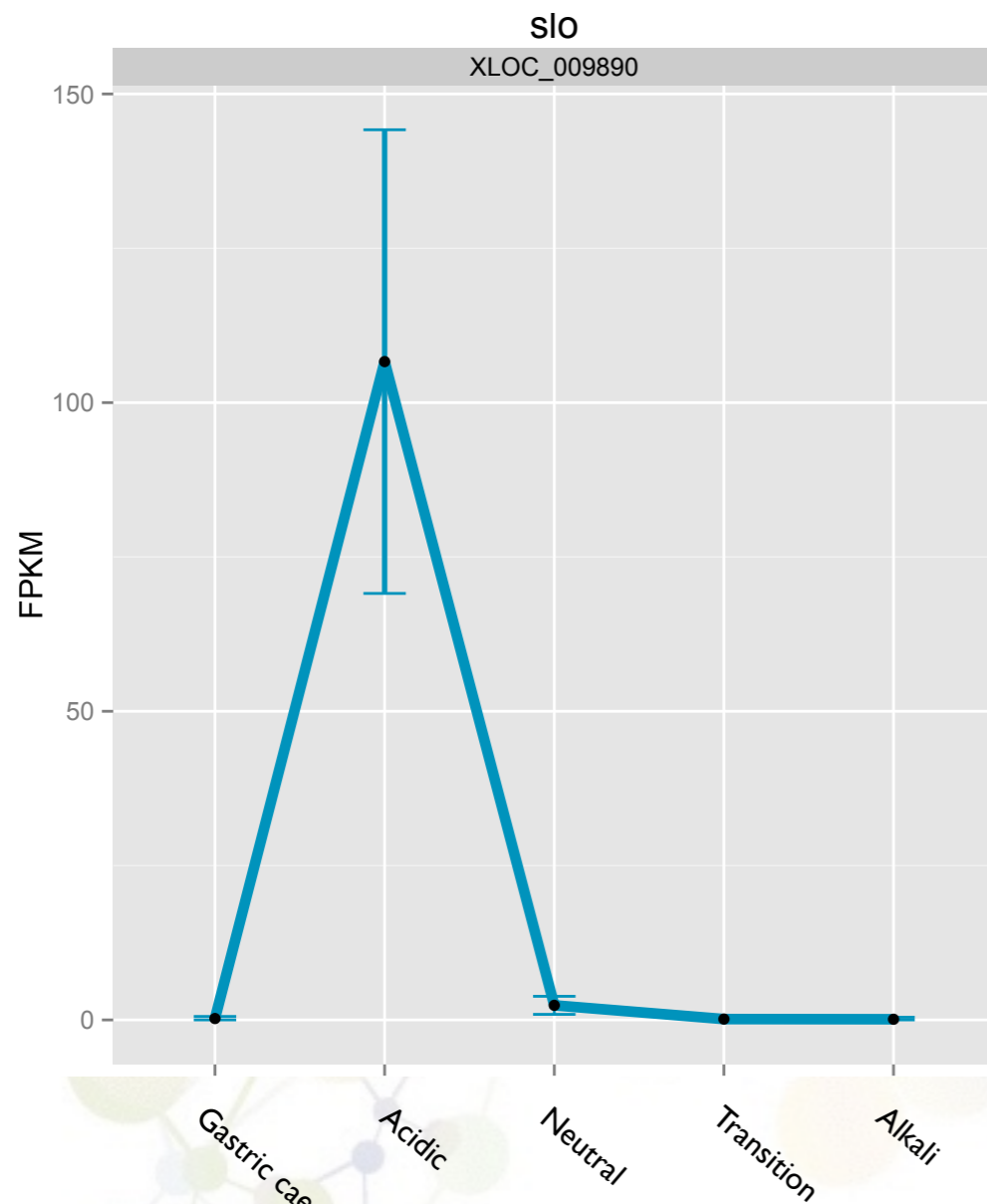
Transcriptomics



Transcriptomics



What does this transcript do?



Control



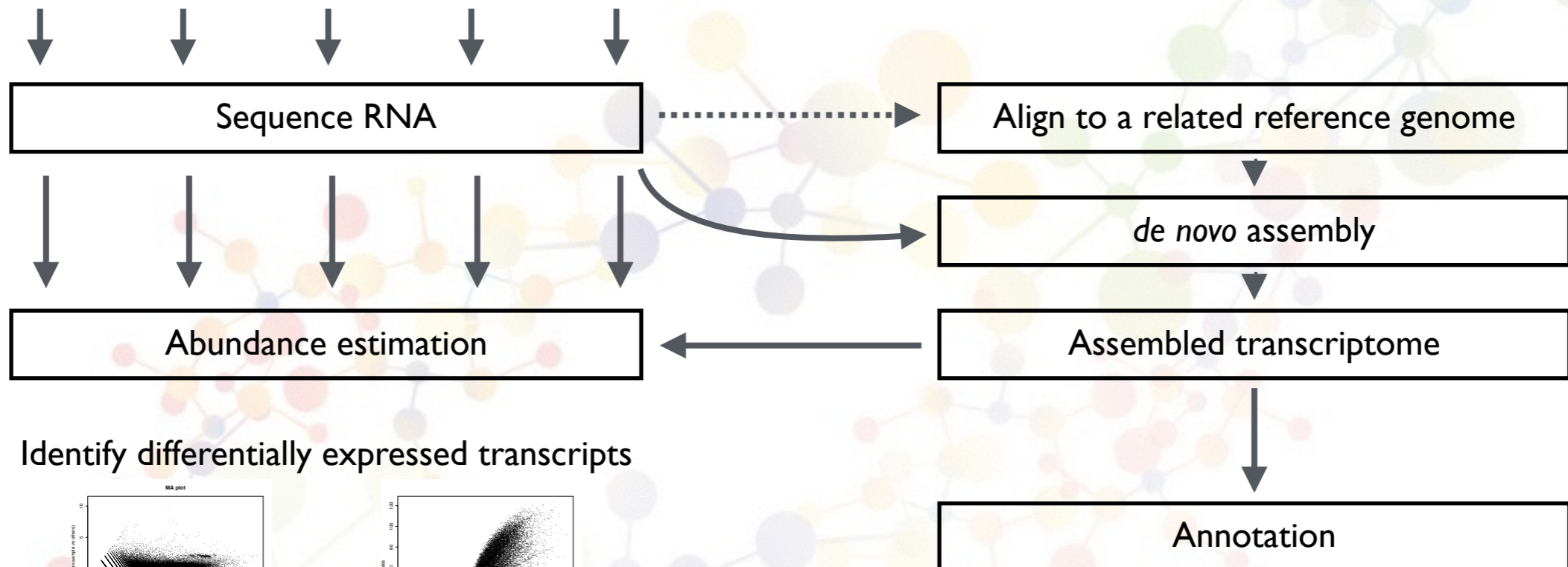
Slowpoke RNAi



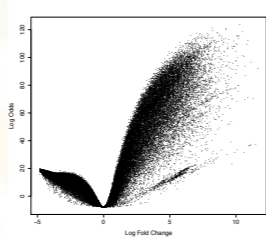
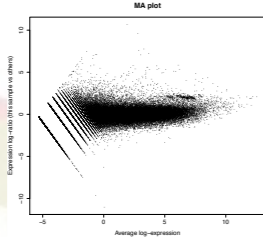
RNA-Seq without a reference



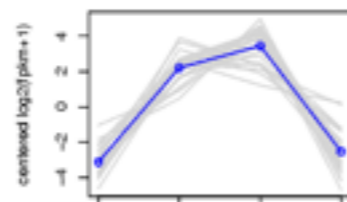
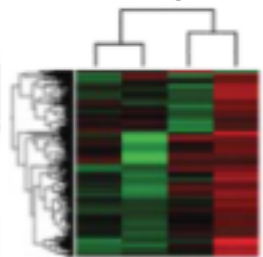
Trinity- Transcriptome assembly



Identify differentially expressed transcripts



Expression patterns, transcript clusters





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